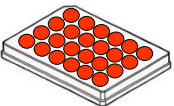
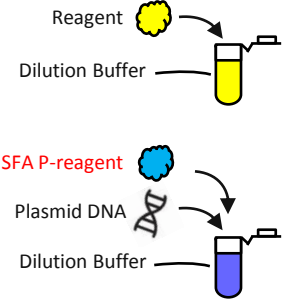

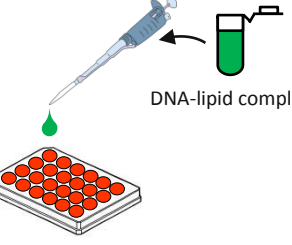
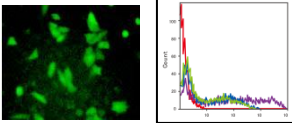


1-Step method (Reverse transfection method)

Timeline	Steps
<p>1</p> <p>Reagent Dilution Buffer</p> <p>SFA P-reagent Plasmid DNA Dilution Buffer</p>	<p>Dilute ScreenFect™ A plus Reagent*¹ in Dilution Buffer, and then mix well *¹ Vortex the reagent before use</p> <p>Dilute DNA and SFA P-reagent in Dilution Buffer, and then mix well</p>
<p>2</p> <p>DNA-lipid complex</p>	<p>Add diluted DNA (+SFA P-reagent) to diluted ScreenFect™ A plus Reagent, and then incubate for 5 minutes ~ at room temperature*² *² Incubation is available until the step 4 has been completed</p>
<p>3</p> <p>Cultured cells</p>	<p>Prepare required cells for transfection</p>
<p>4</p> <p>Cell suspension</p>	<p>Detach cells and prepare the cell suspension, and then transfer the required numbers of cell suspension to cell culture plate</p>
<p>5</p> <p>DNA-lipid complex</p>	<p>Add DNA-lipid complex from step 2 to well of cell culture plate from step 4</p>
<p>6</p> <p>Visualize/analyze transfected cells</p>	<p>Visualize/analyze transfected cells</p>

Procedure Details		96-well	24-well	12-well	6-well
Component					
Dilution Buffer for ScreenFect™ A plus		5 µL	25 µL	50 µL	125 µL
DNA : Transfection Reagent ratio		1 : 3	1 : 4	1 : 3	1 : 4
ScreenFect™ A plus Transfection Reagent		0.15 µL	0.2 µL	0.75 µL	1.0 µL
Dilution Buffer for ScreenFect™ A plus		5 µL	25 µL	50 µL	125 µL
DNA (0.1-2.5 µg / µL)		50 ng	250 ng	500 ng	1250 ng
SFA P-reagent (2µL / µg DNA)		0.1 µL	0.5 µL	1.0 µL	2.5 µL
Diluted DNA (+SFA P-reagent)		5 µL	25 µL	50 µL	125 µL
Diluted ScreenFect™ A plus Transfection Reagent		5 µL	25 µL	50 µL	125 µL
Adherent cells or suspension cells		1.0-4.0 × 10 ⁴	0.5-2.0 × 10 ⁵	1.0-4.0 × 10 ⁵	0.25-1.0 × 10 ⁶
Cell Detachment (Trypsin or Accutase®)					
Final composition [per well]					
DNA-lipid complex		10 µL	50 µL	100 µL	250 µL
DNA amount		50 ng	250 ng	500 ng	1250 ng
SFA P-reagent used		0.1 µL	0.5 µL	1.0 µL	2.5 µL
ScreenFect™ A plus Transfection Reagent used		0.15 or 0.2 µL	0.75 or 1.0 µL	1.5 or 2.0 µL	3.75 or 5.0 µL
Medium volume		100 µL	500 µL	1000 µL	2000 µL
Incubate cells for 1 day ~ at 37°C. Then, analyze transfected cells.					

2-Step method (Forward transfection method)

Timeline		Steps	Procedure Details										
1	Day 0	 Pre-Cultured cells	Seed cells to be 70-90% confluent at transfection	Component									
				Adherent cells or suspension cells		96-well		24-well		12-well		6-well	
			Seed cells to be 70-90% confluent at transfection. The medium replacement may improve a transfection efficiency in a 2-Step method.										
2		 Reagent Dilution Buffer SFA P-reagent Plasmid DNA Dilution Buffer	Dilute ScreenFect™ A plus Reagent*1 in Dilution Buffer, and then mix well ※1 Vortex the reagent before use Dilute DNA and SFA P-reagent in Dilution Buffer, and then mix well	Dilution Buffer for ScreenFect™ A plus									
				DNA : Transfection Reagent ratio		5 µL		25 µL		50 µL		125 µL	
			ScreenFect™ A plus Transfection Reagent										
			Dilution Buffer for ScreenFect™ A plus										
			DNA (0.1-2.5 µg / µL)										
			SFA P-reagent (2µL / µg DNA)										
3	Day 1	 DNA-lipid complex	Add diluted DNA (+SFA P-reagent) to diluted ScreenFect™ A plus Reagent, and then incubate for 5 minutes ~ at room temperature*2 ※2 Incubation is available until the step 4 has been completed	Diluted DNA (+SFA P-reagent)									
				Diluted ScreenFect™ A plus Transfection Reagent		5 µL		25 µL		50 µL		125 µL	
4		 DNA-lipid complex	Add DNA-lipid complex from step 3 to well of cell culture plate from step 1	Final composition [per well]									
				DNA-lipid complex		96-well		24-well		12-well		6-well	
			DNA amount										
			SFA P-reagent used										
			ScreenFect™ A plus Transfection Reagent used										
			Medium volume										
5	Day 2 ~		Visualize/analyze transfected cells	Incubate cells for 1 day ~ at 37°C. Then, analyze transfected cells.									
				For support, please visit the http://screenfect.jp									